

Jan 13, 2025

2. Sample Preparation - An Easy-to-Prepare Sample Slide

 In 1 collection

DOI

dx.doi.org/10.17504/protocols.io.x54v92b9113e/v1



Valeria Berno¹, Kees van der Oord², Luis-Francisco Acevedo-Hueso³, Laszlo Barna⁴, Sebastian Beer⁵, Yann Cesbron⁶, Orestis Faklaris⁷, Nathalie Gaudreault⁸, David Grunwald⁹, Mathias Hammer⁹, Rainer Heintzmann¹⁰, Ayse Aslihan Koksoy¹¹, David McFadden¹², Glyn Nelson¹³, Roland Nitschke¹⁴, Santosh Podder¹⁵, Britta Schroth-Diez¹⁶, Sathya Srinivasan¹⁷, Andre Zeug¹⁸, Gert-Jan Bakker¹⁹

¹ALEMBIC Advanced Light and Electron Microscopy Biolmaging Center, San Raffaele Scientific Institute;

²Nikon Europe BV; ³Neuro-AI Monitoring and Quantum Medical Diagnostics;

⁴Institute of Experimental Medicine, Budapest; ⁵Hamamatsu Photonics Deutschland GmbH;

⁶Institute of Science and Technology Austria;

⁷MRI, Biocampus, University of Montpellier, CNRS, INSERM, Montpellier, France;

⁸Allen Institute for Cell Science, Seattle, WA, USA;

⁹UMass Chan Medical School, RNA Therapeutics Institute, Worcester, MA, USA;

¹⁰Leibniz Institute of Photonic Technology; ¹¹MD Anderson Cancer Center; ¹²Friedrich Schiller University Jena;

¹³Newcastle University, Biolmaging Unit, UK;

¹⁴Life Imaging Center and Signalling Research Centres CIBSS and BIOSO, University of Freiburg, Germany;

¹⁵Indian Institute of Science Education and Research (IISER) Pune;

¹⁶Max Planck Institute of Molecular Cell Biology and Genetics, Dresden, Germany; ¹⁷ONPRC;

¹⁸Cellular Neurophysiology, Hannover Medical School, Hannover, Germany;

¹⁹Radboud University Medical Center, Medical BioSciences department, Nijmegen, the Netherlands

QUAREP-LiMi

Tech. support email: info@quarep.org



Britta Schroth-Diez

Max Planck Institute of Molecular Cell Biology and Genetics,...

OPEN  ACCESS



DOI: dx.doi.org/10.17504/protocols.io.x54v92b9113e/v1

External link: <https://quarep.org/>

Protocol Citation: Valeria Berno, Kees van der Oord, Luis-Francisco Acevedo-Hueso, Laszlo Barna, Sebastian Beer, Yann Cesbron, Orestis Faklaris, Nathalie Gaudreault, David Grunwald, Mathias Hammer, Rainer Heintzmann, Ayse Aslihan Koksoy, David McFadden, Glyn Nelson, Roland Nitschke, Santosh Podder, Britta Schroth-Diez, Sathya Srinivasan, Andre Zeug, Gert-Jan Bakker 2025. 2. Sample Preparation - An Easy-to-Prepare Sample Slide. [protocols.io https://dx.doi.org/10.17504/protocols.io.x54v92b91l3e/v1](https://dx.doi.org/10.17504/protocols.io.x54v92b91l3e/v1)

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

Created: September 08, 2024

Last Modified: January 13, 2025

Protocol Integer ID: 107122

Keywords: ink marker, fluorescent sample, in-homogenous, high dynamic range, marker pen, easy preparation, thin layer

Disclaimer

This protocol collection was developed by members of **WG2 "Detection System Performance"** of the international consortium **QUAREP-LiMi**.

The Consortium for Quality Assessment and Reproducibility for Instruments and Images in Light Microscopy (QUAREP-LiMi), formed by the global community of practitioners, researchers, developers, service providers, funders, publishers, policy makers and industry related to the use of light microscopy, is committed to democratizing access to quantitative and reproducible light microscopy and the data generated by it.

This protocol collection has undergone the internal approval process of QUAREP-LiMi.

It is not the intention of this protocol collection to supplant the need for independent professional judgement, advice, diagnosis or treatment. Any action taken or withheld on the basis of the information presented here is undertaken at the user's own risk. The user agrees that neither the company nor any of the authors, contributors, administrators, or other individuals associated with protocols.io or QUAREP-LiMi can be held liable for any injuries, damages or losses incurred as a result of the user's use of the information contained in or linked to this protocol or any of our websites, applications, or services.

Abstract

This protocol describes how to create a high dynamic range fluorescent sample using basic office and microscopy lab tools. The protocol is based on the usage of the ink of a fluorescent text marker to create a thin fluorescent layer on a microscope cover glass (Olevsko et al., 2021). The result is a carrier with a thin, homogeneous fluorescent film with abrupt edges and a thickness in the order of a few micrometers. This type of sample can be used to make images with a smooth gradient going from maximum intensity to background, as described in the protocols for determination of a detector's photon conversion factor.

Guidelines

Please refer to the introduction protocol for a more detailed description of a detection system, its parameters, and various aims for performance monitoring.

This protocol is part of a collection of protocols developed by QUAREP-LiMi WG2 for characterizing light microscopy detection system performance.

Materials

Yellow fluorescent marker (for instance: Stabilo Boss Original, Art.No70/24, Heroldsberg, Germany)

Note

The marker is a critical component. Different brands have different formulas for the ink, resulting in different layer characteristics after drying of the ink. The Stabilo marker resulted in a 5 μm thin and homogeneous coverage of the glass surface without cracks and granules. Yellow, for excitation within the range of 488 nm.

- Borosilicate microscope cover glass (e.g., 18 x 18 mm², thickness #1.5, Eur. Cat. No. 631-0121, Avantor delivered by VWR, Radnor, Pennsylvania, United States)
- Standard microscope slides (e.g., ground edges frosted, Eur. Cat. No. 631-1553, Avantor delivered by VWR or Microscope slides,CC frosted, Epredia)
- Picodent Twinsil22 (Picodent twinsil®, picodent, Dental Produktions- und Vertriebs GmbH) or Polysil 3481 silicone (Polysil Ltd), for sealing.
- Gloves (vinyl lab gloves)
- EtOH 70% for cleaning microscope cover glass
- Lens paper for cleaning microscope cover glass (e.g., Whatman® lens cleaning tissue, Grade 105, Merck or Linsenpapier, Assistent)
- Permanent marker pen or pencil to write on the microscope slide
- Nail polish



Safety warnings

- ⚠ Ensure you follow general lab safety guidelines for radiation sources and chemicals as outlined within your organisation.

Before start

Collect **all** the materials (see list in Materials tab) and put on the gloves (to avoid contamination of the sample).

Sample preparation

- 1 Fold a sheet of lens paper, put a microscope cover glass in between the folded sheet, wet with ethanol (EtOH, see Materials), and rub to remove contaminations and oily substances.

Note

In case EtOH is not enough to create a clean surface, rub the glass between the fingers with detergent and rinse it with tap water, before cleaning with EtOH again.



Wear gloves when cleaning the coverglass with ethanol and lens paper

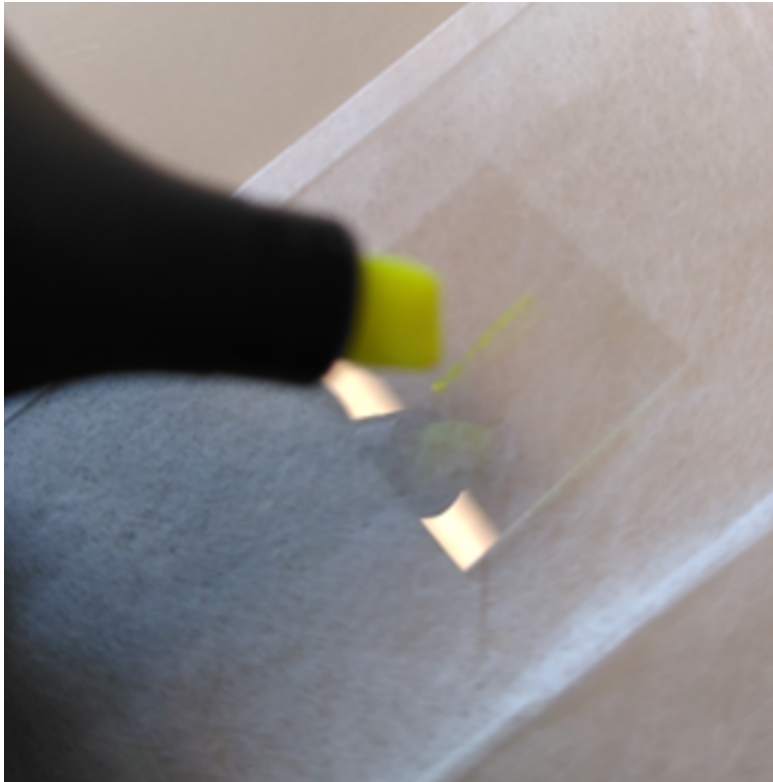
- 2 Dry the cover glass with a fresh sheet of lens paper.

Note

Do this step immediately while the cover glass is still wet. This will ensure that the dissolved contamination is removed. Do not let the contaminants dry onto the cover glass surface again.

- 3 Place the clean cover glass on a fresh sheet of lens paper.

- 4 Draw a thin line on the cover glass with the fluorescent marker using its thin tip.



Drawing a line on cover glass with a marker

- 5 Let the ink on the cover glass dry for 2 minutes.
- 6 Put the cover glass upside down on a microscope slide, with the ink towards the slide.
- 7 Fix the corners of the cover glass on the microscope slide the usual way.

Note

It is not important, what is used to fix the cover slip. Fluorescent marker is not sensitive to nail polish. However, the front lens of high-NA lenses is glued: if nail polish gets on this lens, acetone can seriously damage it. We therefore recommend to use twinsil-type dental silicone to seal the slides. Generally, use only small amounts of the adhesive substance and let it dry long enough.

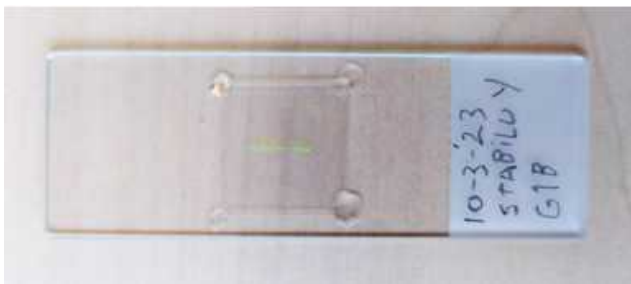
Note

Do not use any embedding medium. It is not necessary for the measurements and might dissolve the marker pen line.

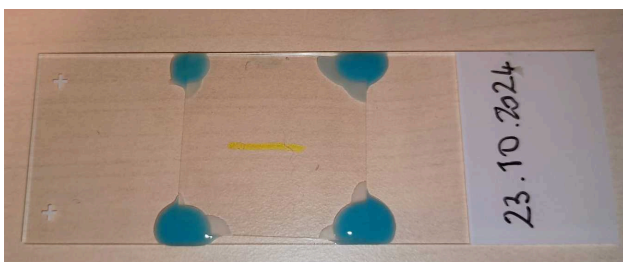
8 Write down the date on the microscope slide.

Note

The marker pen ink will continue to dry out, which might cause cracks and inhomogeneities within part of the ink layer. The sample maybe good to be used for several months, as long as it fullfils the requirements described in the analysis protocol.



End result using nail polish



End result using Twinsil

Protocol references

Olevsko I, Szederkenyi K, Corridon J, Au A, Delhomme B, Bastien T, Fernandes J, Yip C, Oheim M, Salomon A. 2021. A simple, inexpensive and multi-scale 3-D fluorescent test sample for optical sectioning microscopies. *Microscopy Res & Technique* **84**:2625–2635. doi:10.1002/jemt.23813

Acknowledgements

 Funding acknowledgement.pdf 50KB

 CRediT_author_contributions_2_Sa...